Amendments to the claims:

This listing of claims will replace all prior versions, and listings, of claims in the application:

**Listing of Claims:** 

1. (Currently amended) A method for isolating progenitor cells <u>having stem-cell-like</u>

characteristics from a male or female human body, inclusive of all cells with stem-cell-like

characteristics, wherein such cells are isolated directly or indirectly derived from human

mammary secretions comprising, be it colostrum, mature milk, or dry period secretion from

males or females, of said human body during at least one time period selected from the group

consisting of the following periods: a non-pregnant period, a pregnant period, a lactating period,

and an involuting period.

2. (Currently amended) The method according to claim 1, wherein the progenitor cells are

pluripotent or multipotent cells.

3. (Currently amended) A method according to claim 1, wherein said progenitor cells are isolated

from an acellular portion of the mammary secretion in that noncellular parts of the mammary

secretion are that is separated from the a cellular parts portion.

4. (Currently amended) A method according claim 3, wherein non-pluripotent or nonmultipotent

cells are removed from the cellular parts-portion of the mammary secretion.

5. (Currently amended) A method according to any of the preceeding claims claim 1, wherein

human secretory epithelial cells and leucocytes, and microorganisms in particular nonhuman

cells like bacterial cells are removed from the mammary secretion.

6. (Currently amended) A method according to any of the preceding claims claim 1, wherein

progenitor cells are isolated from the-mammary secretions isolated during lactating periods is

used for the isolation of the progenitor cells, and wherein said lactating periods are selected from

the group consisting of the mammary secretion during particular stages of mammary secretion

such as: the period after beginning of individual feeding, and; versus end of individual feeding;

lactation phase; preferably the early lactation period.

7. (Currently amended) A method according to any of the preceding claims <u>claim 1</u>, wherein

magnet beads are used to the isolation of isolate the progenitor cells.

8. (Currently amended) A method according to any of the preceding claims claim 1, wherein in a

first step cellular components are washed out of the mammary secretion, in a second step said

cellular components are stained with antibodies to the progenitor cell markers, and in a third step

the progenitor cells are separated from the other cells directly or indirectly by means of the

attached antibodies.

9. (Currently amended) A method according to claim 8, wherein the antibody-stained progenitor

cells are attached to beads and the progenitor cells are isolated using said beads, wherein when

said beads are preferably small iron beads, said beads are isolated using a magnet, and wherein

the progenitor cells are extracted by means of the beads, preferably in case of small iron beads by

using a magnet, and wherein subsequently the beads or the antibodies or both as well as if-need

be the antibodies are removed from the progenitor cells.

10. (Currently amended) A method according to claim 9, wherein removal of the beads are

removed using an enzyme is effected by means of enzymes selected from the group consisting of

following group: DNase, Proteinase, and RNase.

11. (Currently amended) A method according to any of the preceding claims claim 1, wherein the

progenitor cells are cultured without using a fibroblast feeder layer, in particular without using a

mouse fibroblast feeder layer.

12. (Currently amended) A method according to any of the preceding claims claim 1, wherein in

(i) a first step the whole human mammary secretion is subjected to centrifugation leaving a fat

layer on top, a protein and carbohydrate rich supernatant beneath it, and at the bottom a pellet of

cells;

(ii) in a second step the fat fraction and supernatant are removed;

(iii) in a third step a volume of a buffer or cell culture media, such as, but not limited to,

phosphate buffered saline, tris buffer saline, TBS and/or PBS, or media, such as, but not limited

to, Williams media or RPMI Media, is added and the cells are resuspended in the buffer or media

and centrifuged as in the first step and before, preferentially repeating this step process 3 or 4

times, leaving a substantially pure cell pellet; and

(iv) and in a fourth step separating the progenitor cells are separated from the cell pellet.

13. (Currently amended) A method according to any of the preceding claims claim 12, wherein a

cell pellet is generated from the human mammary secretion, and thereafter subsequently the

following separation steps are used:

(v) the cell pellet is suspended in cell culture media; preferentially in RPMI media containing

foetal-calf (bovine) serum,

(vi) this suspension is incubated for at least 15 minutes at 4°C with progenitor cell-specific or

stem-cell-specific antibodies linked to magnetic beads which have before been incubated with

progenitor, preferentially stem cell-specific antibodies, like antimouse IgG antibodies, which

antibodies are attached to the magnetic beads via a small strand of DNA, wherein the incubation

of the cell suspension is preferentially carried our for 15 minutes at 4°C;

(vii) positioning a magnet in proximity to the suspension, whereby cells that once the progenitor

cells have bound to the magnetic beads a magnet is attached to the tube containing the

cells/beads, thus attracting attract the progenitor cells connected with the beads to the magnet,

whereas unbound cells are not attracted by the magnet and remain in the supernatant; and

(viii) removing the supernatant leaving only the progenitor cells bound to the beads via the

progenitor cell antibody.

14. (Currently amended) A method according to claim 13, wherein thereafter subsequently, the following steps are used:

(ix) progenitor cells bound to the beads via the stem cell-specific antibodies antibody are removed by an appropriate a cleavage means, wherein when preferentially, in case of the antibody being is attached to the beads via small strand of DNA, said cleavage a by means is of addition of a DNase,

(x) the beads are removed by <u>positioning</u> attaching the magnet <u>to attract</u> once more such that the beads, no longer attached to the stem cells, are attracted to it; and

(xi) removing the supernatant now containing the isolated progenitor cells.

15. (Currently amended) A method according to any of the preceding claims claim 1, wherein the cells are separated from human mammary secretion by centrifugation, and subsequently incubated in a growth media that is permissive for growth of progenitor cells, stem cells or lactocyte growth.

16. (Currently amended) A method according to claim 15, wherein in

(i) a first step the <u>unfractionated</u> whole-human mammary secretion is subjected to centrifugation leaving a fat layer on top, a protein and carbohydrate rich supernatant beneath it, and at the bottom a pellet of cells;

(ii) in a second step, the cell pellet is washed in <u>cell culture</u> media;, preferably in RPMI media only

(iii) in a third step the cells <u>comprising</u> of the cell pellet are plated onto a cell culture vessel treated device in bacteriocidal, <u>andlor</u> fungicidal <u>or both bacteriocidal and fungicidal growth media</u> and <u>incubated for no less than 10 and no more than 30 days and thereafter are allowed to incubate, preferably for 10-30 days, most preferably for 14-20 days,</u>

(iv) the cells are harvested, preferably by trypsination, and washed, preferably using buffer or growth media, and

(v) the harvested cells are plated onto a reconstituted basement membrane preparation for growth preferably up to confluence.

17. (Currently amended) A method according to claim 16, wherein in step (v) a—the solubulized basement membrane preparation is extracted from EHS mouse sarcoma is-used, as e.g. MatrigelTM.

18. (Currently amended) Progenitor cells, preferentially pluripotent Pluripotent or multipotent progenitor cells, derived using a method according to any of the preceding claims claim 1.

19. (Currently amended) A method for creating cells or tissues in a mother or infant comprising administering to the mother or infant Use of pluripotent or multipotent progenitor cells prepared according to the method of claim 1 as derived using a method according to any of the claims 1-17 for ex vivo, in vitro and/or in vivo applications.

20. (Cancelled) A use according to claim 19, to create tissues or cells for the benefit of the mother and/or of the infant and/or of other individuals.

21. (Currently amended) A method according to claim 19, further comprising Use according to claim 17 or 20, including subsequent-gene therapy treatments or intrauterine foetal treatments.

22. (Currently amended) A method according to claim 19, wherein the cells or tissues are administered Use according to claims 19-21, for the generation of 20 cells, tissue, glands or organs for the treatment of disease.

23. (Cancelled) Use according to any of the claims 19-23 for subsequent cloning or scientific research.

24. (Currently amended) A method of claim 19, wherein the cells or tissues are administered for Use according to any of the claims 19-23, for one or several of the group of the following purposes: clinical, diagnostic, diagnosis, bioengineering, lactoengineering, breast tissue regeneration, breast reconstructive surgery, breast cosmetic or enhancement surgery, exocrine gland tissue regeneration and/or surgery.

Please enter the above amendments.

Respectfully submitted,

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